

# Entomopathogenic fungi to suppress BMSB

## Participants:

Thomas Pike (Graduate Student), Paula Shrewsbury, Ray St. Leger,  
Department of Entomology, UMD



## Funding



United States  
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Agriculture

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Specialty Crop Research Initiative  
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## Collaborating Institutions



Cornell University



Virginia Tech



# Fungi to suppress BMSB

## **BMSB SCRI Grant (3 year, 2014)**

**Subobjective 2.2.2 - Identification of a single, well characterized strain of fungi expressing the optimum toxin combination for BMSB control**

**Subobjective 2.2.3 - Develop attract-and-kill and mass trapping strategies for management of BMSB in commercial crops**

## **BMSB SCRI Renewal Grant**

**Subobjective 2.2.2 - Development of BMSB-Specific Fungal Entomopathogens (Ray St. Leger, Department of Entomology, UMD)**



# Entomopathogenic Fungi

- ▶ Fungi that colonize and kill their insect host
- ▶ Effective at controlling insects that won't readily consume topical pathogens
  - *Bacillus thuringiensis* (Bt)
- ▶ *Metarhizium anisopliae*, *Beauveria bassiana* and *Isaria fumosorosea* among the most common entomopathogenic fungi used
- ▶ Used in insect systems such as grasshoppers, flies, beetles, caterpillars and greenhouse pests



# Objectives

- ▶ Determine the effects of entomopathogenic fungi on brown marmorated stink bug
  - Wild-type fungal strains
  - Additives (diatomaceous earth and horticultural oil)
  - Transgenic fungal strains
- ▶ Explore their potential as biological control agents



# Wild-type Fungal Bioassay

## ▶ Strains evaluated:

- *Metarhizium* (M): ARSEF 1548, ARSEF 2547, ARSEF 1055, F52 (4 strains)
- *Beauveria* (B): GHA, Botanigard (2 strains)
- *Isaria* (I): ARSEF 3581 (1 strain)
- Other (U): ARSEF 10386, Unidentified fungus (isolated from lab colony) (2 strains)



# Wild-type Fungal Bioassay Methods

- ▶ Fungi plated on PDA media
- ▶ Spore suspensions of  $1 \times 10^7$  conidia/mL used with .01% Tween (later DI water)
- ▶ BMSB submerged in suspension, placed in plastic bowls with food and water
- ▶ Stink bugs monitored for mortality
- ▶ Tested on nymphs and adults
- ▶ Not all treatments represented in each bioassay (9 bioassays)



# Wild-Type Bioassays

Bioassay #	Treatment	Mean % Mortality			
		x % Mortality (Day 3)		x % Mortality (Day 7)	
		Adults	Nymphs	Adults	Nymphs
1	1548 (M)	0	3.33 (±3.33)	10 (±4.47)	30 (±4.47)a
1	2547 (M)	3.33 (±3.33)	0	13.33 (±6.67)	43.33 (±9.54)a
1	Tween	3.33 (±3.33)	0	3.33 (±3.33)	0b
1	Water	3.33 (±3.33)	0	10 (±4.47)	0b
2	1055 (M)	10 (±10)	46.67 (±15.20)	36.67 (±12.01)	100 (±0)
2	GHA - Botanigard (B)	3.33 (±3.33)	40 (±7.30)	30 (±11.25)	90 (±6.83)
2	Tween	10 (±6.83)	40 (±13.66)	40 (±15.49)	80 (±7.30)
2	Water	6.67 (±4.21)	26.67 (±8.43)	33.33 (±9.88)	73.33 (±9.88)
3	2575 (M)	3.33 (±3.33)	40 (±7.30)	43.33 (±8.02)	93.33 (±4.21)
3	Unidentified Fungus (U)	10 (±6.83)	20 (±7.30)	56.67 (±9.54)	93.33 (±4.21)
3	Tween	10 (±4.47)	16.67 (±6.14)	66.67 (±8.43)	86.67 (±6.67)
3	Water	0	16.67 (±8.02)	33.33 (±8.43)	83.33 (±6.14)
4	F52 (M)	25 (±5.00)	25 (±12.58)	60 (±8.16)	70 (±12.91)
4	GHA - USDA (B)	30 (±12.91)	20 (±14.14)	65 (±9.57)	65 (±9.57)
4	3581 (I)	20 (±8.16)	40 (±14.14)	55 (±5.00)	80 (±0)
4	Water	25 (±18.93)	35 (±9.57)	45 (±18.93)	65 (±5.00)
5	10386 (U)	-	13.33 (±6.67)b	-	33.33 (±8.43)b
5	3581 (I)	-	30 (±8.56)a	-	73.33 (±8.43)a
5	Tween	-	23.33 (±3.33)b	-	33.33 (±6.67)b
5	Water	-	3.33 (±3.33)b	-	13.33 (±6.67)b

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5	Tween	-	23.33 (±3.33)b	-	33.33 (±6.67)b
5	Water	-	3.33 (±3.33)b	-	13.33 (±6.67)b

# Wild-Type Fungal Bioassays with Additives

- ▶ Diatomaceous earth (DE) and horticultural oil tested in conjunction with F52 (M), GHA (B) and 3581 (I)
- ▶ Testing for increased mortality with additives
- ▶ 50g/L DE, 7% oil used in treatments
- ▶ All other procedures as before



# Wild-Type Bioassays with Additives

Treatment	Mortality	
	x % Mortality (Day 3)	x % Mortality (Day 7)
Water	5.00 (±5.00)	30.00 (±12.91)
DE	20.00 (±8.16)	35.00 (±9.57)
DE/Oil	40.00 (±11.54)*	60.00 (±8.16)
Oil	40.00 (±8.16)*	55.00 (±12.58)
3581 (I)	20.00 (±0)	45.00 (±9.57)
3581/DE	5.00 (±5.00)	40.00 (±14.14)
3581/DE/Oil	20.00 (±0)	50.00 (±10.00)
3581/Oil	50.00 (±10.00)*	65.00 (±9.57)
F52 (M)	5.00 (±5.00)	35.00 (±5.00)
F52/DE	0 (±0)	50.00 (±10.00)
F52/DE/Oil	10.00 (±10.00)	65.00 (±12.58)
F52/Oil	40.00 (±21.60)*	70.00 (±10.00)
GHA (B)	5.00 (±5.00)	10.00 (±5.77)
GHA/DE	0 (±0)	25.00 (±15.00)
GHA/DE/Oil	20.00 (±8.16)	35.00 (±9.57)
GHA/Oil	15.00 (±5.00)	30.00 (±10.00)

# Wild-Type Bioassays with Additives

Treatment	Mortality	
	x % Mortality (Day 3)	x % Mortality (Day 7)
Water	5.00 (±5.00)	30.00 (±12.91)
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DE/Oil	40.00 (±11.54)*	60.00 (±8.16)
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3581/DE	5.00 (±5.00)	40.00 (±14.14)
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3581/Oil	50.00 (±10.00)*	65.00 (±9.57)
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F52/DE	0 (±0)	50.00 (±10.00)
F52/DE/Oil	10.00 (±10.00)	65.00 (±12.58)
F52/Oil	40.00 (±21.60)*	70.00 (±10.00)
GHA (B)	5.00 (±5.00)	10.00 (±5.77)
GHA/DE	0 (±0)	25.00 (±15.00)
GHA/DE/Oil	20.00 (±8.16)	35.00 (±9.57)
GHA/Oil	15.00 (±5.00)	30.00 (±10.00)

# Transgenic Fungal Bioassays

- ▶ Use of transgenic fungi successful in other insect systems
  - Scorpion and spider toxins
- ▶ Fungi engineered to express spider neuropeptides
- ▶ *Metarhizium* strains tested: Hv1a-1548, Dc1a-1548, As1a-1548, Ta1a-1548
- ▶ All other procedures as before



# Transgenic Bioassays

Treatment	Mortality		Fungal Growth	
	$\bar{x}$ % Mortality (Day 3)	$\bar{x}$ % Mortality (Day 7)	$\bar{x}$ Days to Growth	$\bar{x}$ % dead with Growth
1548 WT	20 ( $\pm 14.14$ )	95 ( $\pm 5.00$ )	4 ( $\pm 0.87$ )	40 ( $\pm 8.16$ )
As1a	10 ( $\pm 5.77$ )	80 ( $\pm 14.14$ )	2 ( $\pm 0$ )	20 ( $\pm 8.16$ )
Dc1a	15 ( $\pm 9.57$ )	70 ( $\pm 10.00$ )	2.67 ( $\pm 1.11$ )	40 ( $\pm 8.16$ )
Hv1a	30 ( $\pm 10.00$ )	65 ( $\pm 5.00$ )	3.25 ( $\pm 1.43$ )	40 ( $\pm 8.16$ )
Ta1a	20 ( $\pm 8.16$ )	75 ( $\pm 9.57$ )	8 ( $\pm 0$ )	10 ( $\pm 5.77$ )
Water	20 ( $\pm 8.16$ )	60 ( $\pm 14.14$ )	-	-



# Summary

- ▶ Overall levels of mortality low
- ▶ Little difference seen between wild-type strains (with and without additives) and transgenic strains
- ▶ Low virulence of fungi
- ▶ No indication that any of the entomopathogenic fungi evaluated would be effective as a biological control ☹️



# Mechanism behind low virulence of fungi against brown marmorated stink bug

- ▶ Many insects protect themselves from fungal infection via chemical defense
  - Earwigs
  - Sawflies
  - Bed bugs
  - Pentatomids
- ▶ Brown marmorated stink bug defensive compounds may be the cause of poor fungal performance





# Objectives

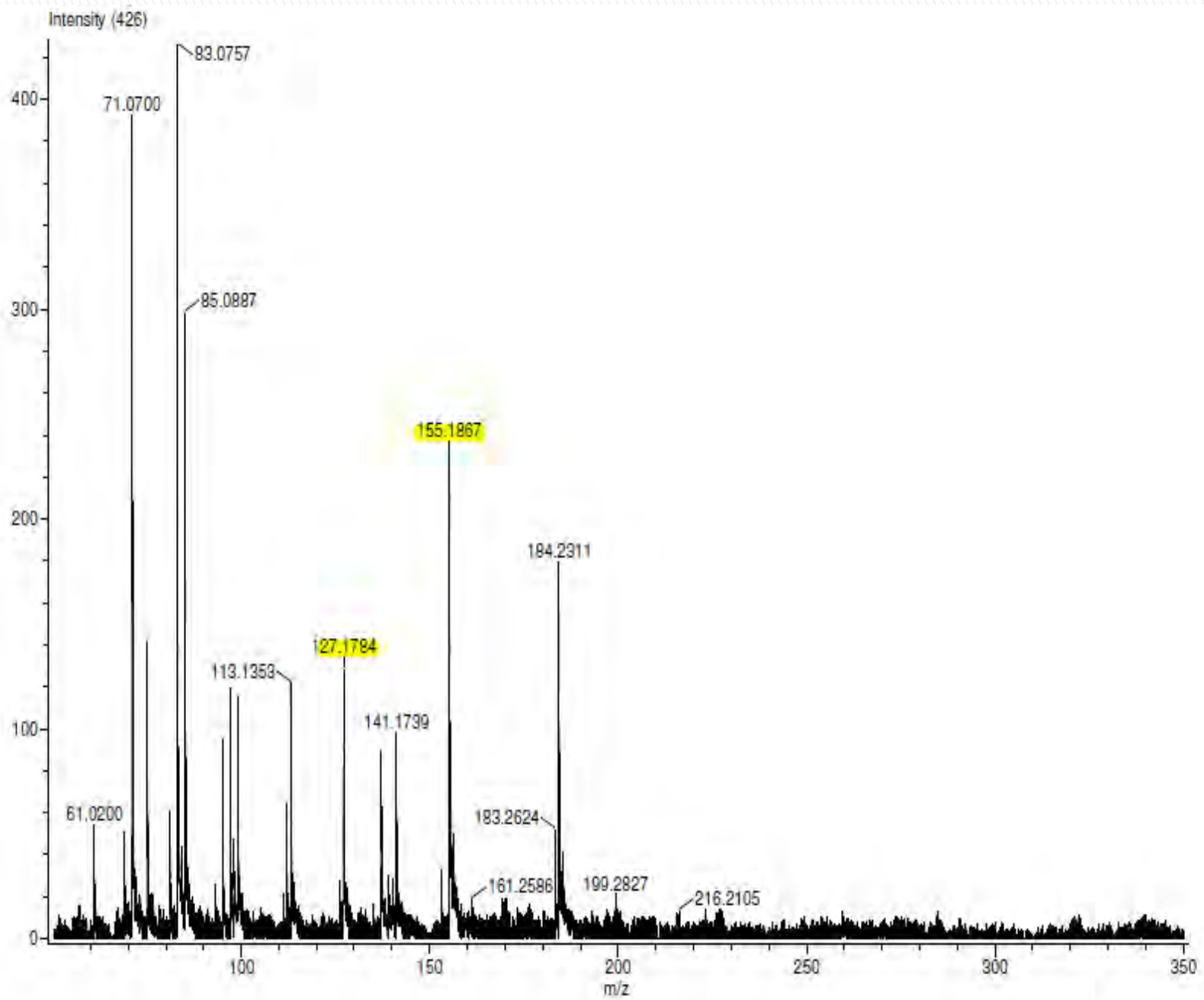
- ▶ Identify constituents of brown marmorated stink bug defensive compound
- ▶ Evaluate effects of defensive compounds against entomopathogenic fungi
  - Fungal growth
  - Spore germination



# Defensive compounds

- ▶ From literature and other studies
  - Too many chemicals to ID, based search on *a priori* hypothesis
  - Predict that trans-2-octenal and trans-2-decenal are potential candidates
- ▶ Analyze brown marmorated stink bug secretions to confirm presence of trans-2-octenal and trans-2-decenal
  - AccuTOF mass spectrometer equipped with confined Direct Analysis in Real Time (cDART) ion source





# Defensive compound effect on fungal growth – Inhibition

- ▶ 3 fungi evaluated
  - F52 (*Metarhizium*), GHA (*Beauveria*), ARSEF 3581 (*Isaria*)
- ▶ Fungi plated on PDA media
- ▶ 5  $\mu$ L trans-2-octenal or trans-2-decenal applied to filter paper disc on inside lid of petri dish
  - 100%, 10%, 1%, 0% concentrations



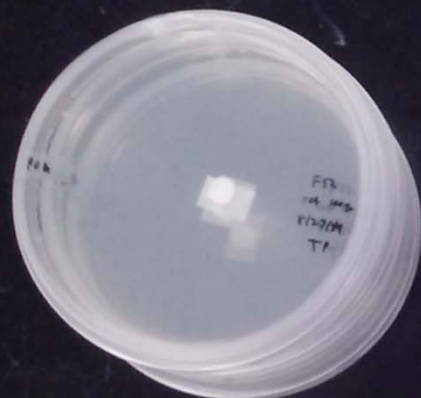
# Defensive compound effect on fungal growth – Inhibition

- ▶ Inhibition of fungi
  - Fungi grown for 3 days at 27° C
  - # of plates with / without fungal growth recorded

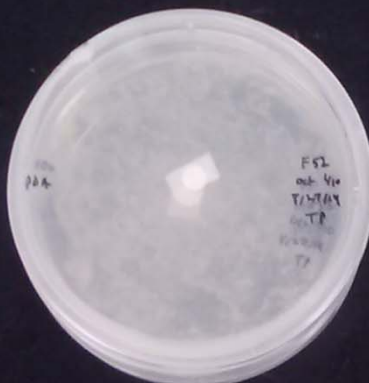


# Trans-2-octenal

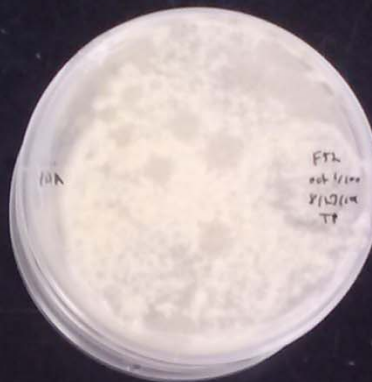
100%



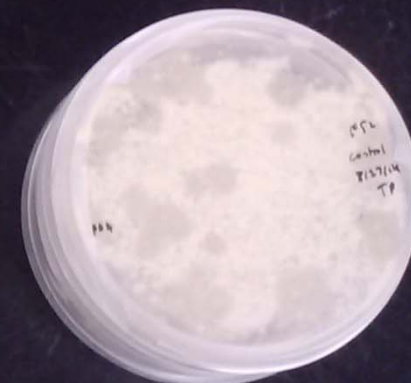
10%



1%

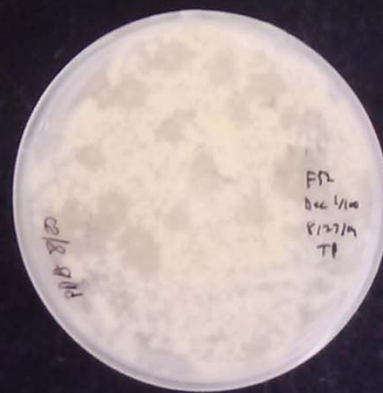
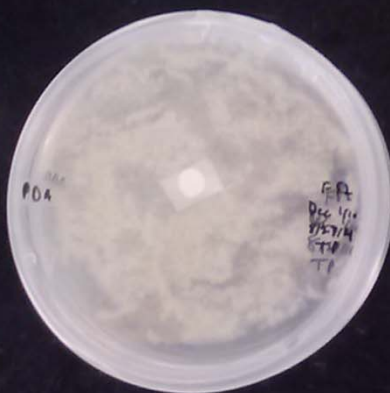


0%



Control

# Trans-2-decenal



Fungus	Mean % Petri dishes without Fungal Growth (Inhibition)						
	Trans-2-octenal Concentration			Trans-2-decenal Concentration			Control
	100%	10%	1%	100%	10%	1%	
F52 (M)	100% ( $\pm 0$ )	66% ( $\pm 33.33$ )	0 ( $\pm 0$ )	100% ( $\pm 0$ )	33% ( $\pm 33.33$ )	0 ( $\pm 0$ )	0 ( $\pm 0$ )
GHA (B)	100% ( $\pm 0$ )	100% ( $\pm 0$ )	0 ( $\pm 0$ )	100% ( $\pm 0$ )	66% ( $\pm 33.33$ )	0 ( $\pm 0$ )	0 ( $\pm 0$ )
3581 (I)	100% ( $\pm 0$ )	0 ( $\pm 0$ )	0 ( $\pm 0$ )	100% ( $\pm 0$ )	33% ( $\pm 33.33$ )	0 ( $\pm 0$ )	0 ( $\pm 0$ )

- ▶ Complete inhibition of fungal growth at 100% concentration
- ▶ No inhibition at 1% or 0% concentrations
- ▶ Partial inhibition at 10% concentration for both chemicals, and all fungal strains



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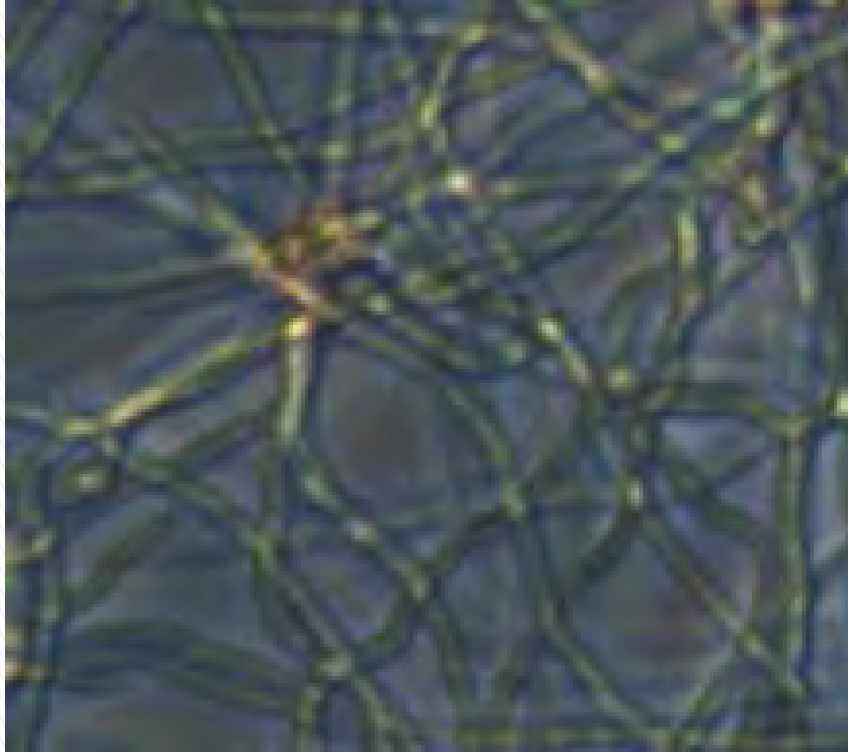
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# Defensive compound effect on spore germination

- ▶ 2 mL liquid PDB media containing spore suspension added to 35 mm petri dishes
- ▶ Trans-2-octenal and trans-2-decenal added at same concentrations as previous experiment
- ▶ Dishes photographed at 400x magnification 1 day after treatment, % spore germination calculated
- ▶ Spore germination defined as presence of germ tube

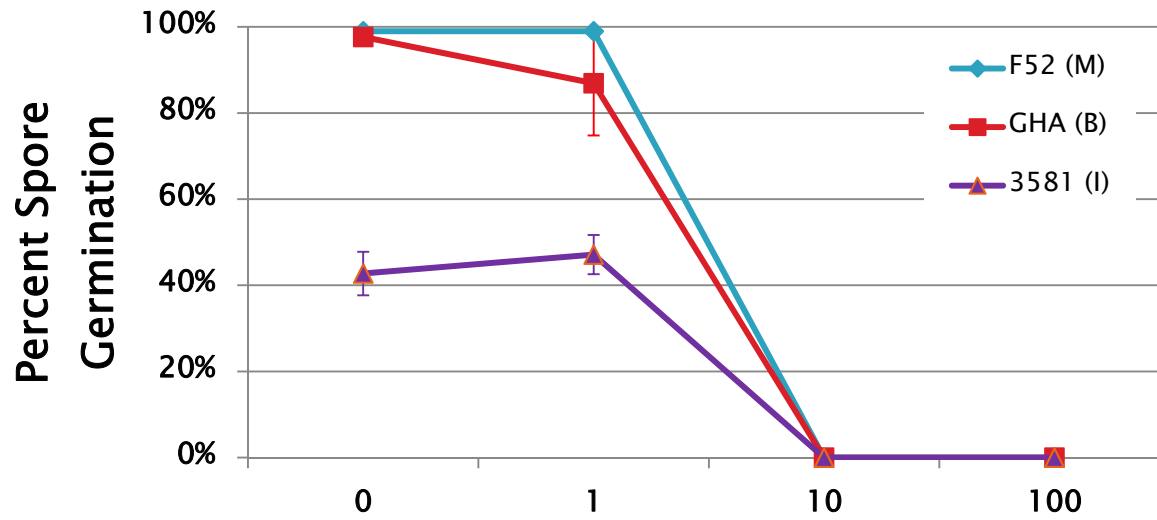




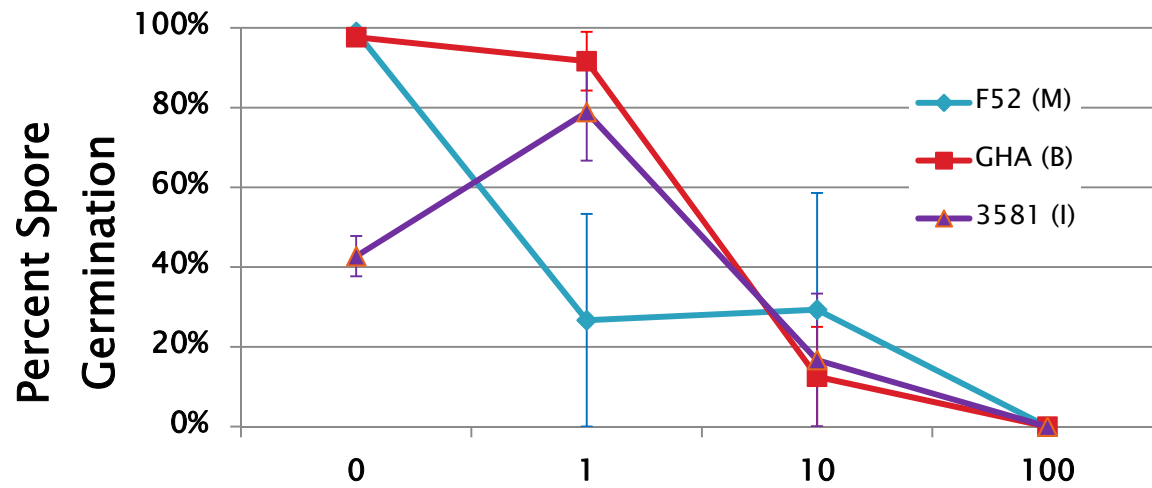
**Germinated F52**



**Ungerminated F52**



Trans-2-octenal Concentration (%)



Trans-2-decenal Concentration (%)



# Summary

- ▶ **Trans-2-octenal and trans-2-decenal present in live brown marmorated stink bug defensive secretions**
- ▶ **Both chemicals inhibit fungal growth and spore germination in laboratory setting**



# Conclusions

- ▶ **Additional challenges with using entomopathogenic fungi as biological control**
  - Low and inconsistent mortality
  - Additives and transgenic strains do not improve efficacy
- ▶ **Defensive chemicals may explain low virulence of fungi against brown marmorated stink bug**
  - Inhibition of fungal growth/spore germination



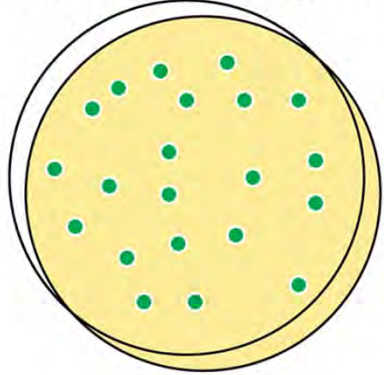
# Creating a pseudo-wild type strain

- ▶ Inducing mutations in wild-type strains with the hopes of getting a beneficial mutation
- ▶ Attempt to induce mutation that confers resistance to stink bug defensive compounds
- ▶ Fewer obstacles to use than with GMO products





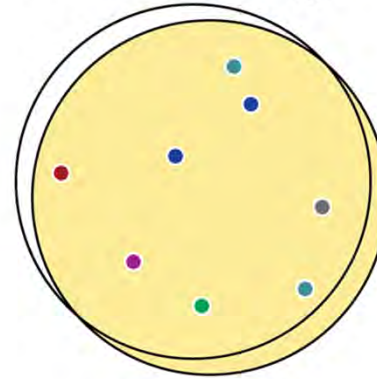
Stink Bug  
Specific Fungus



UV-C Exposure



UV-Generated  
Mutant Fungi

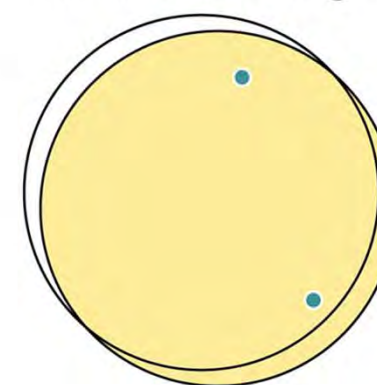


# UV Mutagenesis Methodology

BMSB Volatile  
Exposure



Volatile Resistant  
Mutant Fungi



Bioassays With  
Volatile Resistant  
Mutants



# Acknowledgements

## Thesis Committee

Paula Shrewsbury  
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Joseph Patt

Nancy Harding  
Galen Dively Lab  
Ray St. Leger Lab  
Hsiao-Ling Lu  
Brian Lovett  
Yue Li  
Joe Torella



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